# An Automated Liquid Handling and Sample Preparation Workflow for LC-MS/MS Analysis of Vitamin D Metabolites, Testosterone, and Cortisol

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### **Key Words**

Versette, ControlMate, Prelude SPLC, SRM, bioanalytical validation

### Goal

To develop an automated research workflow that eliminates manual pipetting from the analysis of vitamin D metabolites, testosterone, and cortisol; and to evaluate the accuracy and precision of data acquired using the method.



Figure 1. The Versette automated liquid handler (left) and 96-channel head with Thermo Scientific<sup>™</sup> D.A.R.T.'S<sup>™</sup> tips (right) used for the preparation of vitamin D, testosterone, and cortisol

### **Experimental**

The Versette automated liquid handler was used to aliquot samples, internal standards, and protein precipitation reagents. A Thermo Scientific<sup>™</sup> Prelude SPLC<sup>™</sup> system was equipped with Thermo Scientific<sup>™</sup> TurboFlow<sup>™</sup> Cyclone-P<sup>™</sup> column (50 x 0.5 mm) and a Thermo Scientific<sup>™</sup> Accucore<sup>™</sup> PFP analytical column (50 x 2.1 mm, 2.6 µm particle size). The detector for the system was a Thermo Scientific<sup>™</sup> TSQ Vantage<sup>™</sup> triple-stage quadrupole mass spectrometer equipped with a HESI-II ion source.

### Sample Preparation

Samples of 25-hydroxyvitamin  $D_2$ , 25-hydroxyvitamin  $D_3$ , and testosterone were prepared in human serum. The cortisol sample was prepared in urine. The optimum sample preparation protocol for each method was created and designed using the Thermo Scientific<sup>TM</sup> Versette<sup>TM</sup> ControlMate<sup>TM</sup> software (shown in Figure 2).

A glass or plastic sample-containing plate was placed onto the Versette stage. Then, 100  $\mu$ L of each sample vial was aliquoted into a 2 mL, 96-well plate. To each sample, 200  $\mu$ L of crashing reagent containing internal standard was added. The samples were vortexed for 30 s and centrifuged at 4500 rpm for 3 min. Then, 200  $\mu$ L of supernatant was transferred into a clean plate for injection.



## Introduction

The use of liquid chromatography-tandem mass spectrometry (LC-MS/MS) for the analysis of vitamin D metabolites, testosterone, and cortisol is well established. This note presents a method for analysis of these compounds that incorporates automated liquid handling and eliminates manual pipetting through the use of a Thermo Scientific<sup>TM</sup> Versette<sup>TM</sup> automated liquid handler (Figure 1) for sample preparation. The accuracy and precision of the method were tested using the analysis of 25-hydroxyvitamin D<sub>2</sub>, 25-hydroxyvitamin D<sub>3</sub>, testosterone, and cortisol. For validation, the data generated was required to fall within a set of parameters outlined by Thermo Scientific standard operating procedures (SOPs). These parameters include, but are not limited to:

- The lower limit of quantitation (LLOQ) and low quality control (LQC) must be ±20% of the expected concentration.
- All of the remaining calibrators and controls must be ±15%.
- To determine both inter- and intraday accuracy and precision, these requirements must be met for three consecutive days.



Figure 2. Versette ControlMate software

### Liquid Chromatography

LC system	Prelude SPLC system powered by TurboFlow technology
Mobile phases	A: aqueous phase, 10 mM ammonium formate, 0.01% formic acid in water B: organic phase, 10 mM ammonium formate, 0.01% formic acid in methanol C: column wash, 45% isopropyl alcohol, 45% acetonitrile and 10% acetone <i>All mobile phase solvents were Fisher Chemical brand.</i>
Needle washes	Aqueous: 60% water, 40% methanol, and 0.5% formic acid Organic: 45% isopropyl alcohol, 45% acetonitrile and 10% acetone
Columns	TurboFlow Cyclone-P column (50 x 0.5 mm) Accucore PFP analytical column (50 x 2.1 mm, 2.6 µm particle size) encased in a 70 °C column heater

## Mass Spectrometry

The mass spectrometer was a TSQ Vantage triple-stage quadrupole mass spectrometer equipped with a heated electrospray ionization (HESI-II) probe. It was operated in positive ion, selected-reaction monitoring (SRM) mode.

## Results and Discussion Vitamin D

The analytical measurement range for 25-hydroxyvitamin  $D_2$ and 25-hydroxyvitamin  $D_3$  was 2.0–100 ng/mL. The MRM chromatograms at the LLOQ and the calibration curves for 25-hydroxyvitamin  $D_2$  and  $D_3$  are shown in Figures 3 and 4, respectively. The accuracy and precision data for the quality controls for the three days of validation for 25-hydroxyvitamin  $D_2$  and  $D_3$  are listed in Tables 1 and 2, respectively. The coefficient of variability ( $R^2$ ) for all three days for both analytes ranged from 0.9918 to 0.9980.

## Testosterone

The analytical measurement range for testosterone was 0.020–10 ng/mL The SRM chromatogram at the LLOQ and the calibration curve for testosterone are shown in Figure 5. The accuracy and precision data for the quality controls for the three days of validation for testosterone are listed in Table 3. The R<sup>2</sup> values for all three days ranged from 0.9978 to 0.9985.

## Cortisol

The analytical measurement range for cortisol was 3.62-362 ng/mL. The SRM chromatogram at the LLOQ and the calibration curve for cortisol are shown in Figure 6. The accuracy and precision data for the quality controls for the three days of validation for cortisol are listed in Table 4. The R<sup>2</sup> values for all three days ranged from 0.9971 to 0.9988.



Figure 3. SRM chromatogram of 25-hydroxyvitamin  $D_2$  at the LLOQ and an example calibration curve



Low QC	Day 1		Day 2		Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
6.0 6.0 6.0 6.0 6.0	6.26 6.73 5.50 5.46 6.00	4.3 12.1 -8.4 -9.1 -0.1	4.93 5.46 6.01 6.77 6.43	-17.8 -9.1 0.1 12.9 7.2	6.79 5.18 5.30 6.12 5.03	13.2 -13.6 -11.7 2.0 -16.2
Average SD %RSD	5.99 0.53 8.9	-0.2	5.92 0.74 12.5	-1.4	5.68 0.75 13.2	-5.3
Mid QC	Da	y 1	Da	y 2	Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
40.0 40.0 40.0 40.0 40.0	34.9 38.7 36.5 35.0 40.4	-12.8 -3.2 -8.7 -12.4 0.9	38.7 41.2 39.4 39.2 36.7	-3.4 3.1 -1.5 -2.0 -8.2	37.0 37.0 38.5 37.9 39.6	-7.5 -7.6 -3.7 -5.2 -1.0
Average SD %RSD	37.1 2.38 6.4	-7.2	39.1 1.62 4.1	-2.4	38.0 1.11 2.9	-5.0
High QC	Day 1		Day 2		Day 3	
Concentration (ng/mL)	Concentration (ng/mL)	%Difference	Concentration (ng/mL)	%Difference	Concentration (ng/mL)	%Difference
80.0 80.0 80.0 80.0 80.0 Average	72.3 75.6 74.7 78.6 82.5 76.7	-9.6 -5.5 -6.6 -1.8 3.1 -4.1	85.1 80.7 81.9 81.7 84.0 82.7	6.4 0.9 2.3 2.1 5.0 3.3	80.3 77.9 80.5 85.6 80.8 81.0	0.4 -2.7 0.7 7.0 1.0 1.3
SD %RSD	3.91 5.1		1.81 2.2		2.82 3.5	





Figure 4. SRM chromatogram of 25-hydroxyvitamin  $D_3$  at the LLOQ and an example calibration curve

Table 2. Accuracy and precision data for 25-hydroxyvitamin  $\rm D_{_3}$ 

Low QC	Day 1		Day 2		Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
6.0 6.0 6.0 6.0 6.0	6.24 6.00 5.49 5.98 5.35	4.1 -0.1 -8.5 -0.4 -10.9	6.30 5.83 5.81 6.77 5.63	4.9 -2.8 -3.2 12.9 -6.3	6.09 6.16 6.65 6.65 6.49	1.4 2.7 10.9 10.8 8.1
Average SD %RSD	5.81 0.38 6.54	-3.1	6.07 0.47 7.74	1.1	6.41 0.27 4.21	6.8
Mid QC	Da	y 1	Da	y 2	Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
40.0 40.0 40.0 40.0 40.0	35.6 38.6 39.0 36.0 41.2	-11.00 -3.462 -2.607 -9.97 2.92	41.9 40.3 37.7 38.8 38.7	4.81 0.7 -5.65 -3.08 -3.14	39.2 37.4 39.0 40.8 38.1	-2.04 -6.50 -2.57 1.97 -4.79
Average SD %RSD	38.1 2.29 6.01	-4.8	39.5 1.64 4.15	-1.3	38.9 1.28 3.29	-2.8
High QC	Da	y 1	Day 2		Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
80.0 80.0 80.0 80.0 80.0 Average	78.5 83.5 79.2 82.0 80.3 80.7	-1.8 4.4 -1.0 2.5 0.4 0.9	81.2 79.2 79.3 77.9 78.2 79.2	1.5 -1.0 -0.9 -2.7 -2.2 -1.1	81.7 81.4 77.7 80.9 84.1 81.2	2.1 1.7 -2.9 1.1 5.2 1.4
SD %RSD	2.04 2.5		1.28 1.62		2.32 2.86	



Figure 5. SRM chromatogram of testosterone at the LLOQ and an example calibration curve

Table 3. Accuracy and pre	ecision data for testosterone
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Low QC	Day 1		Day 2		Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
0.06 0.06 0.06 0.06 0.06 0.06	0.064 0.058 0.061 0.063 0.059	6.18 -3.44 1.84 4.91 -1.18	0.054 0.063 0.058 0.064 0.059	-10.06 5.50 -3.95 5.99 -1.30	0.054 0.061 0.058 0.057 0.062	-10.42 1.13 -2.52 -5.31 3.40
Average SD %RSD	0.061 0.002 3.28	1.66	0.059 0.004 6.72	-2.45	0.059 0.003 5.09	-2.74
Mid QC	Da	y 1	Da	y 2	Da	y 3
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
0.45 0.45 0.45 0.45 0.45	0.438 0.450 0.468 0.461 0.435	-2.58 0.05 4.05 2.54 -3.36	0.434 0.422 0.456 0.439 0.451	-3.53 -6.23 1.33 -2.34 0.23	0.413 0.435 0.422 0.436 0.415	-8.20 -3.32 -6.23 -3.06 -7.87
Average SD %RSD	38.1 2.29 6.01	0.14	0.44 0.014 3.18	-2.11	0.42 0.011 2.59	-5.74
High QC	Da	y 1	Day 2		Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
8 8 8 8 8	8.04 7.88 7.89 7.95 7.84	0.54 -1.49 -1.41 -0.68 -2.00	7.89 7.84 8.07 8.04 7.96	-1.35 -2.02 0.91 0.55 -0.46	7.85 8.18 8.12 8.18 7.98	-1.82 2.24 1.44 2.25 -0.28
Average SD %RSD	7.92 0.079 1.00	-1.01	7.96 0.099 1.24	-0.47	8.06 0.142 1.76	0.77





Figure 6. SRM chromatogram of cortisol at the LLOQ and an example calibration curve

Low QC	Day 1		Day 2		Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
10.0 10.0 10.0 10.0 10.0	10.9 10.2 9.5 9.7 n/a	9.3 2.3 -4.7 -3.4 n/a	9.3 9.5 9.8 9.6 10.7	-7.1 -5.5 -1.7 -3.6 7.2	10.4 9.8 10.6 11.0 10.6	3.5 -1.6 6.2 9.6 6.1
Average SD %RSD	10.09 0.64 6.34	0.9	9.79 0.56 5.72	-2.1	10.48 0.42 4.01	4.8
Mid QC	Da	y 1	Da	y 2	Day 3	
Theoretical Concentration (ng/mL)	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference	Measured Concentration (ng/mL)	%Difference
145.0 145.0 145.0 145.0 145.0 145.0	136 138 130 137 134	-6.52 -4.70 -10.25 -5.50 -7.70	132 128 138 141 132	-8.67 -12.0 -4.94 -2.98 -9.14	141 144 148 140 144	-2.93 -0.64 2.25 -3.43 -0.76
Average SD %RSD	134.9 3.14 2.33	-6.9	134.1 5.17 3.86	-7.5	143.4 3.27 2.28	-1.1
High QC	Da	y 1	Da	y 2	Da	y 3
Concentration (ng/mL)	Concentration (ng/mL)	%Difference	Concentration (ng/mL)	%Difference	Concentration (ng/mL)	%Difference
290.0 290.0 290.0 290.0 290.0	278 298 317 311 310	-4.1 2.7 9.5 7.3 6.8	286 NA 275 291 300	-1.4 36.4 -5.3 0.3 3.5	313 318 334 315 317	7.9 9.5 15.1 8.7 9.3
Average SD %RSD	302.8 15.54 5.10	4.4	288.0 10.40 0.04	6.7	319.3 8.34 2.61	10.1

## Conclusion

- The Versette automated liquid handler provided accuracy and precision with high efficiency. All methods were validated, with the R<sup>2</sup> values ranging from 0.9918 to 0.9988.
- The Versette automated liquid handler demonstrated 90% time savings over manual pipetting and passed all acceptance criteria for the compounds tested. Approximate sample preparation time for one full 96 well plate for these methods, when performed manually, ranges from 2 to 3 hours; whereas the Versette automated liquid handler sample preparation time was 10–15 minutes. This minimized technician sample handling and saved time, money, and resources.
- The Versette system, in combination with the Prelude SPLC system and the TSQ Vantage mass spectrometer, allowed for highly automated preparation of samples for LC-MS/MS research.

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